

# Differentiation of Enantiomers in Complex Systems by NMR Spectroscopy and Chiral Solvating Agents (CSA) : Applications and Methodology



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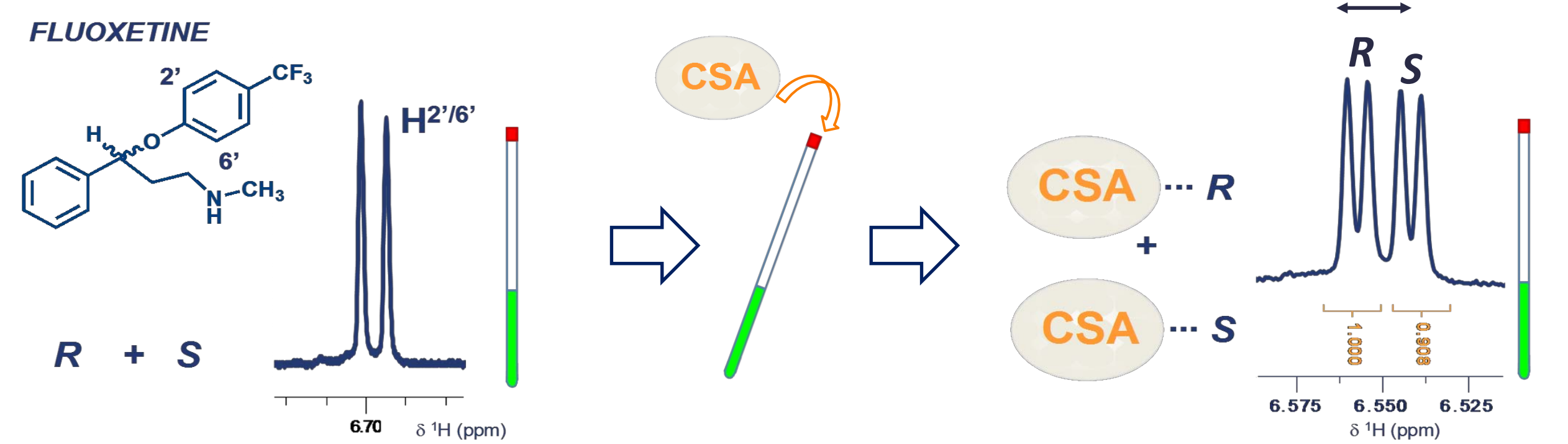
## INTRODUCTION

### Complex Systems

- ✓ Mixture of compounds:
  - body fluids (urine, plasma, ...)
  - extracts of plants/tissues
  - crude of reactions
- ✓ Pure enantiomeric mixtures with complex  $^1\text{H}$  spectrum

### Enantiodifferentiation by CSA & NMR Spectroscopy

Example : pure sample in an organic solvent by  $^1\text{H}$  NMR <sup>1</sup>



Enantiodifferentiation of racemic fluoxetine in  $\text{CDCl}_3$  using ABTE-18 as CSA

### Advantages

- ✓ Simple
- ✓ Fast
- ✓ Robust
- ✓ No derivatization
- ✓ No purification

### Fields of Application

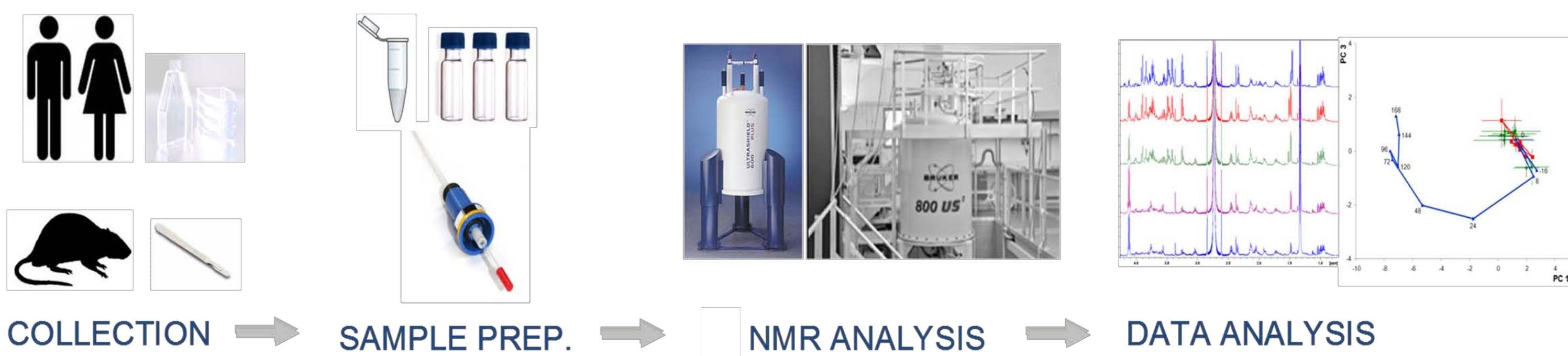
- ✓ Pharmacology
- ✓ Chiral Metabonomics <sup>2</sup>
- ✓ Natural Products
- ✓ Toxicity Studies
- ✓ ...

## CHIRAL METABONOMICS <sup>2</sup> - NEW APPLICATION

### Towards Enantiospecific Metabolic Profiling

**Metabolic Profile:** “description of the small molecule composition of a biological sample”

#### Outline of Analysis Strategy



Most of endog./exog. metabolites are chiral molecules

Does not differentiate enantiomers of chiral metabolites

BLIND TO POTENTIAL ENANTIOMERIC RATIO DIFFERENCES

### Enantiodifferentiation of *R,S*-ibuprofen within human urine by $^1\text{H}$ NMR and a CSA

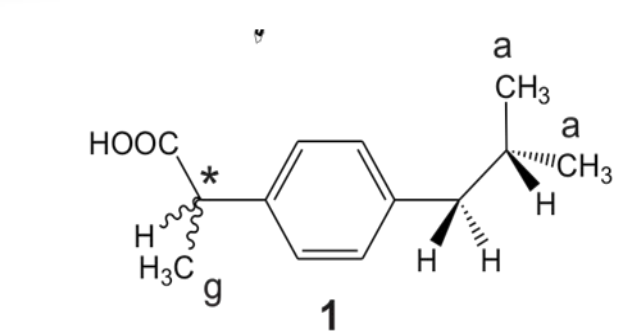
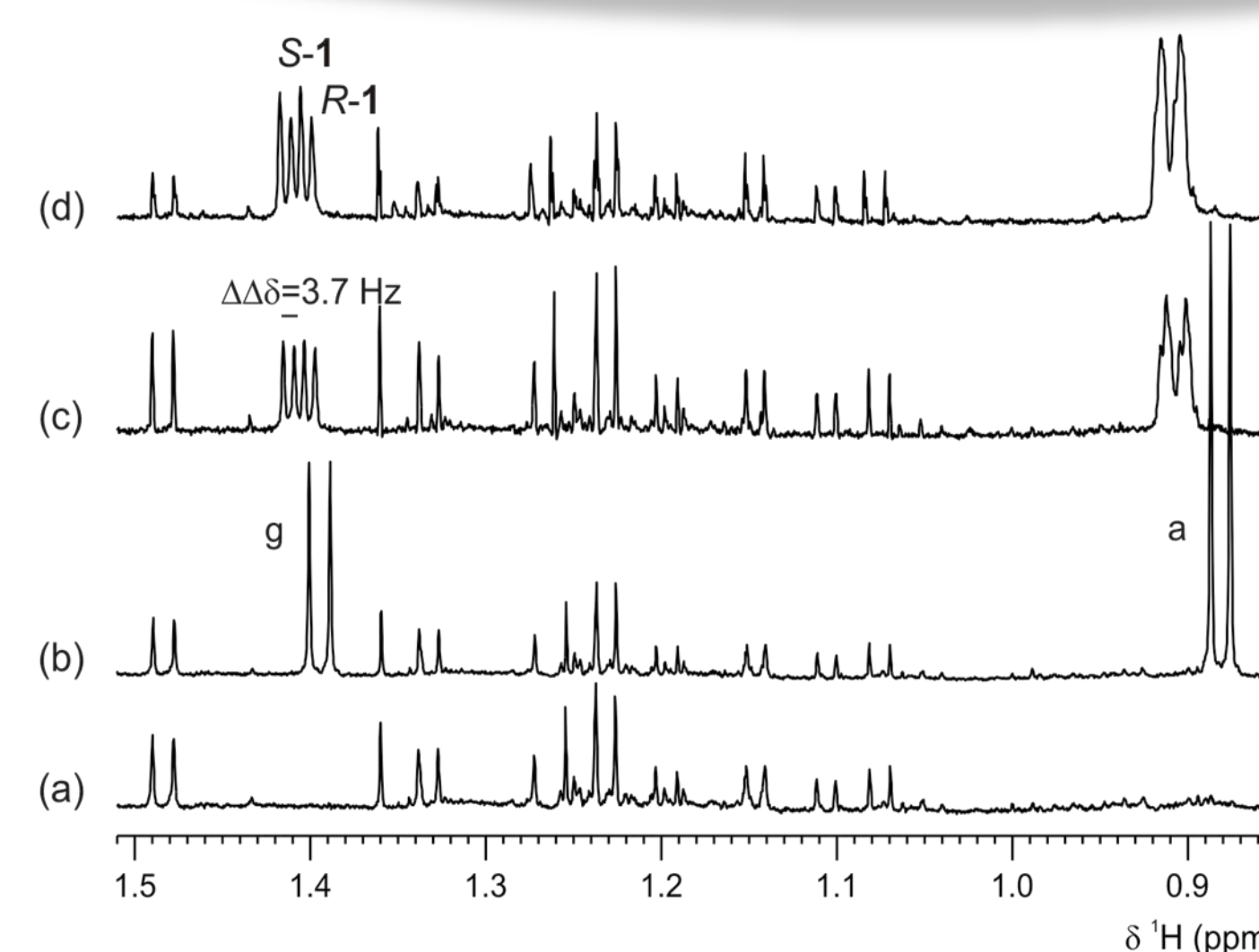


Figure 1. Expanded region of  $^1\text{H}$  NMR spectra of (a) human urine of a healthy volunteer, (b) after the addition of racemic ibuprofen **1**, (c) the former sample in presence of  $\beta\text{CD}$ , (d) after spiking sample (c) with *S*-1. Spectra were acquired at 300.0 K at a magnetic field strength of 14.1 T.

## $^{13}\text{C}$ NMR <sup>3</sup>: OVERCOME THE OVERLAPPING PROBLEM - METHODOLOGY

### Enantiodifferentiation by $^{13}\text{C}$ NMR & CSAs

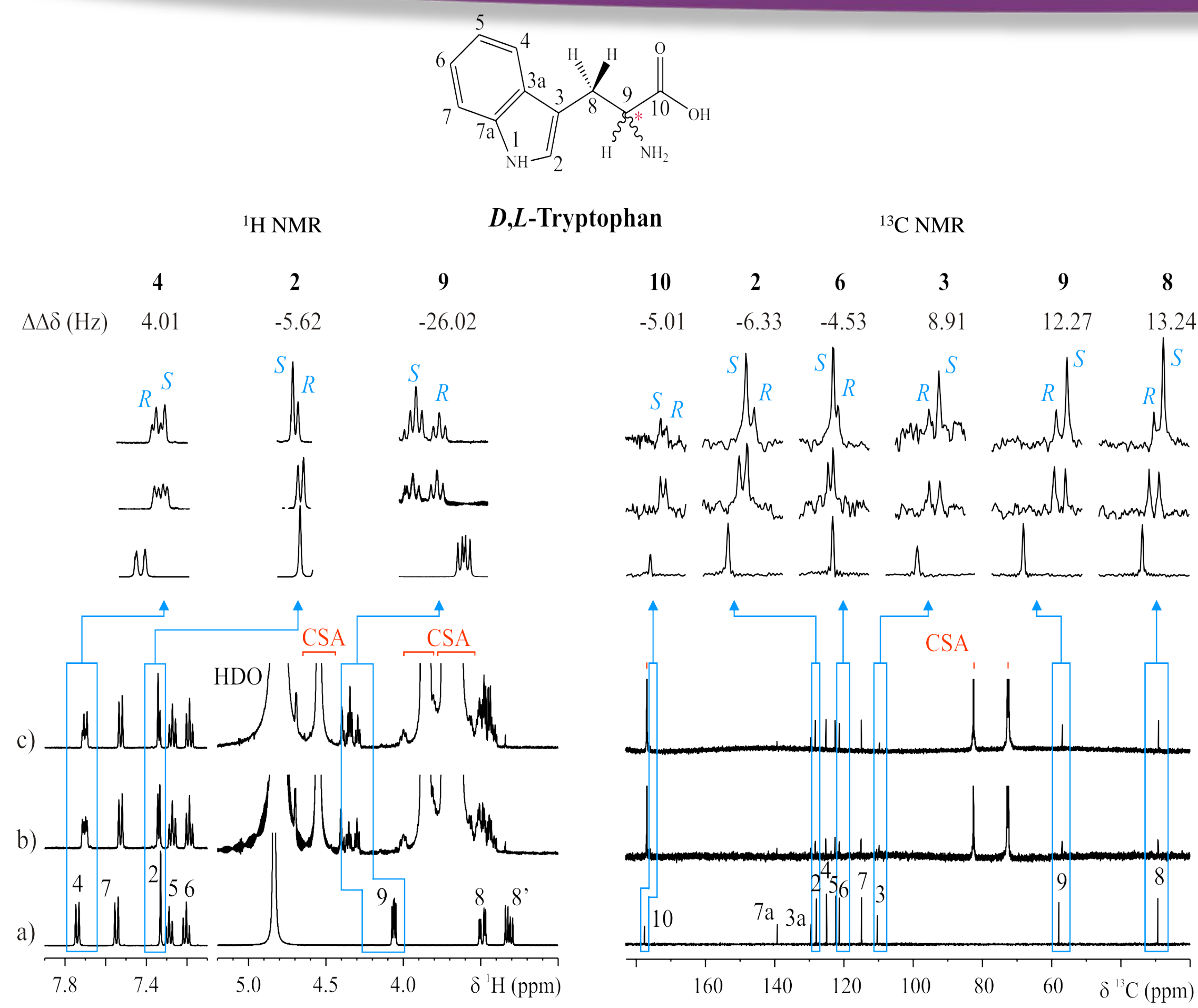


Figure 2. a) 2.3 mM Racemic TRP in  $\text{D}_2\text{O}$ ; b) with 5.4 equivalents of 18C6TCA added and c) after spiking the sample with L-TRP. Experiments performed in a 500 MHz spectrometer equipped with a TCI cryoprobe.

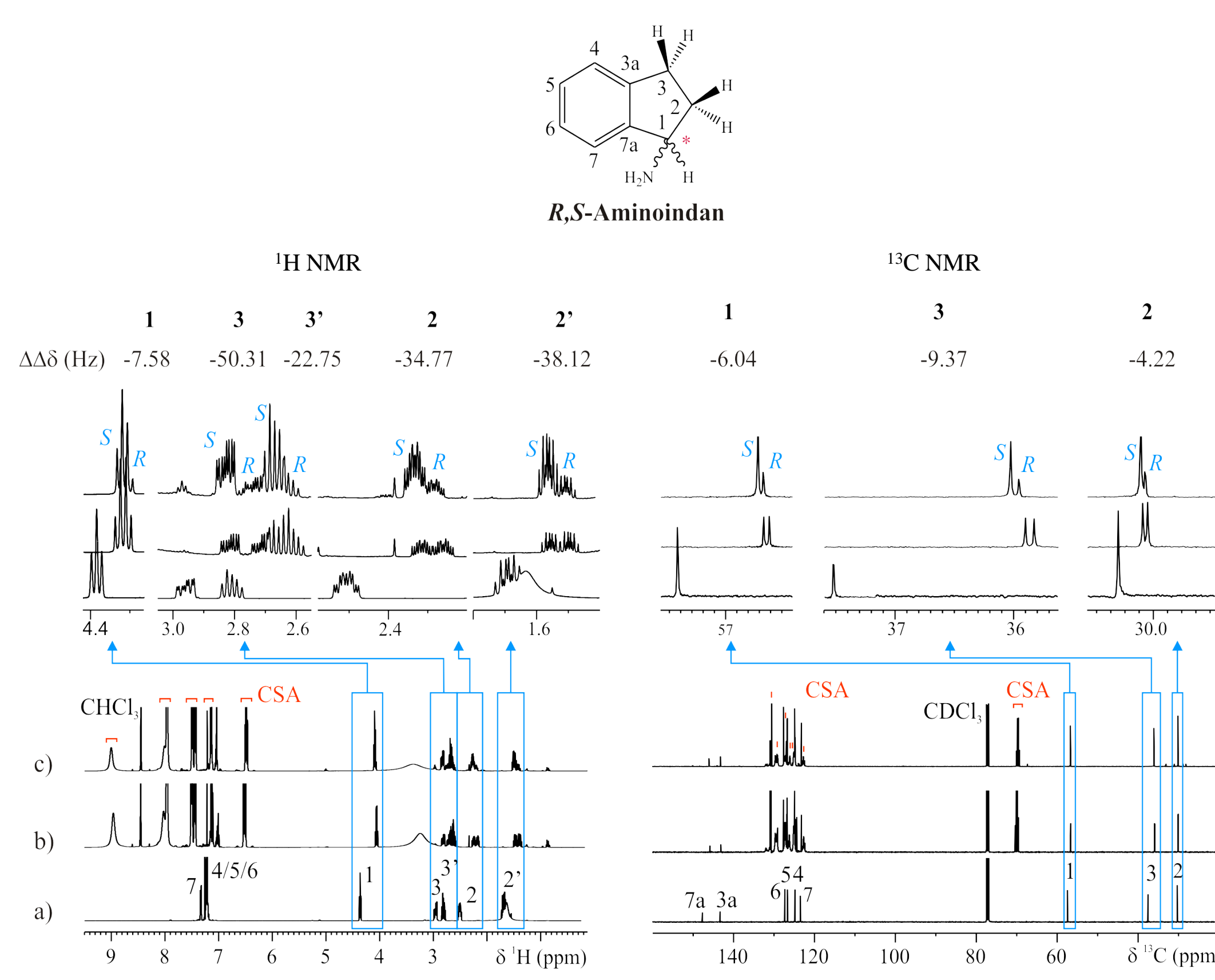


Figure 3. a) 50 mM Racemic AMI in  $\text{CDCl}_3$ ; b) with 4.5 equivalents of *R*-PA added and c) after spiking the sample with *S*-AMI. Experiments performed in a 500 MHz spectrometer equipped with a TCI cryoprobe.

### *R/S* Molar Ratio Measurement

Table 1. Theoretical and measured (by the ratio of the signal integrals) *R/S* molar ratio values of mixtures of *R,S*-AMI. Measured values correspond to three different experiments: 1D  $^1\text{H}$  (zg, r.d. 2s), 1D  $^{13}\text{C}$  with NOE contribution (zgpg, r.d. 2 s) and 1D  $^{13}\text{C}$  without NOE contribution (zgig, r.d. 8 s). The observational error for each measurement in percentage is indicated. The three mixtures were prepared from a 50 mM racemic AMI solution and the CSA used was 4,5 equivalents of *R*-PA.

Theoretical <i>S/R</i> ratio <sup>a</sup> ( <i>R/S</i> )	Measured <i>R/S</i> ratio and Error <sup>b</sup>							
	$^1\text{H}$				$^{13}\text{C}$			
	Hatom	meas.	error (%)		Catom	meas.	error (%)	
1 (50:50)	H-1	0.98	1.62 *		C-1	1.01	1.25	1.00 0.11
	H-2	1.01	1.28 *		C-2	1.00	0.35	1.01 0.99
	H-2'	0.97	2.92 *		C-3	1.00	0.31	0.99 0.71
	H-3	0.77	22.90 **p		C-3a	0.98	2.40	1.03 2.66
	H-3'	-	-	**/**				
3 (25:75)	H-1	3.08	2.50 *		C-1	3.00	0.14	3.00 0.03
	H-2	4.34	44.57 *		C-2	3.00	0.05	3.03 1.00
	H-2'	3.38	12.73 *		C-3	3.06	1.98	3.02 0.62
	H-3	-	-	**	C-3a	2.91	2.87	3.10 3.20
	H-3'	-	-	**/**				
9 (10:90)	H-1	6.98	22.44 *		C-1	9.05	0.57	8.81 2.10
	H-2	11.11	23.41 *		C-2	8.92	0.90	8.95 0.58
	H-2'	10.11	12.30 *		C-3	8.97	0.29	9.01 0.11
	H-3	-	-	**	C-3a	-	-	-
	H-3'	12.58	39.80 *					

<sup>a</sup> From weighted values

<sup>b</sup> Observational error (fm-fr)\*100/fr

\*Signal partially split

\*\*p) Signal (partially) overlapped with another signal of the spectrum

## CONCLUSIONS

There is an interest in the differentiation (and identification) of enantiomers in complex systems, such as complex mixtures. This is the case of enantiodifferentiation studies of biological samples. Recently, a new area in metabonomics was described named under “Chiral Metabonomics”, in which enantiomeric molecules are differentiated in a metabolic profile; the experiment was conducted using a CSA and  $^1\text{H}$  NMR spectroscopy.<sup>2</sup>

The differentiation of enantiomers in complex samples by  $^1\text{H}$  NMR is often impeded by overlapping, due to the small  $\delta$  range of proton and to the multiplicity of the signals. Though experimental times are longer than for  $^1\text{H}$ , observing  $^{13}\text{C}$  nuclei is a convenient information-rich alternative in many situations and particularly when studying complex systems, since 1D  $\{^1\text{H}\}$   $^{13}\text{C}$  NMR experiment:

- overcomes the main overlapping drawback of the  $^1\text{H}$  NMR experiment due to the intrinsic high dispersion of the  $^{13}\text{C}$  and to the easiness of obtaining a proton decoupled spectrum.
- provides valuable/complementary information to the  $^1\text{H}$  NMR experiment and extends the possibilities of enantiodifferentiation to fully deuterated and nonproton containing compounds.
- has a huge potential for the enantiomeric study of complex mixtures (Chiral Metabonomics).



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<sup>1</sup> Pérez-Trujillo, M.; Virgili, A., Tetrahedron-Asymmetr. **2006**, 17, 2842-2846. <sup>2</sup> Pérez-Trujillo, M.; Lindon, J.C.; Parella, T.; Keun, H.; Nicholson, J.K.; Athersuch, T.J. Anal. Chem. **2012**, 84, 2868-2874. <sup>3</sup> Pérez-Trujillo, M.; Monteagudo, E.; Parella, T. **2013 Submitted**.

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